# SquiggleFilter

#### An Accelerator for Portable Virus Detection

Tim Dunn, Harisankar Sadasivan, Jack Wadden, Kush Goliya, Kuan-Yu Chen, David Blaauw, Reetuparna Das, and Satish Narayanasamy University of Michgan MICRO '21

Presented by: Joël Lindegger joel.lindegger@inf.ethz.ch 23/12/2021





### **Executive Summary**

**Motivation:** Portable nanopore sequencers enable **virus detection early** in a pandemic, before PCR or antigen tests are available

**Problem:** - Sequencing output must be labeled as "viral" or "non-viral" in real-time

- Existing solutions are too slow by ≥2x and will not scale for future sequencers

- Most of the work done by existing solutions is unnecessary

Goal: - Accelerate the labelling process to keep up even with future sequencers

**Challenge:** - Basecalling is a necessary step in the pipeline and the main **bottleneck** 

- It is data-dependent, tools may become invalid every 6 months, thus people are

discouraged to improve it

**Key Ideas:** - Filter data before the basecalling step (in raw signal space)

- accelerate the filter in hardware

**Results:** - 274x more throughput

- 2x less power usage

(baseline: Guppy-lite on NVIDIA Jetson AGX Xavier)





#### Outline

- 1. Background and Motivation
  - 1. Virus detection
  - 2. Nanopore sequencing
- 2. SquiggleFilter
  - 1. Pipeline
  - 2. Accelerator
- 3. Results
- 4. Strengths/Weaknesses
- 5. Discussion

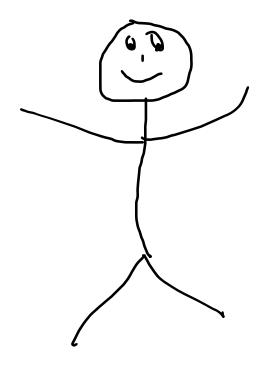


#### Outline

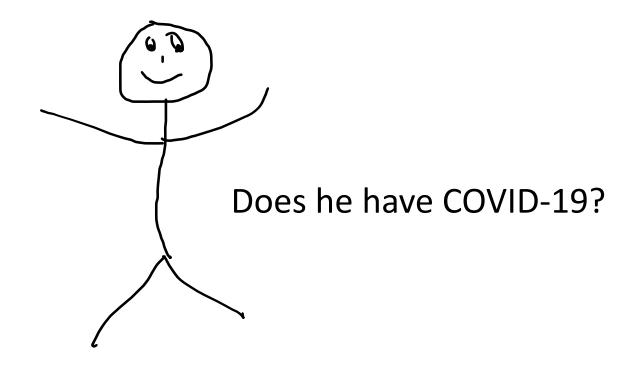
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Non-Programmable

Programmable





#### Non-Programmable



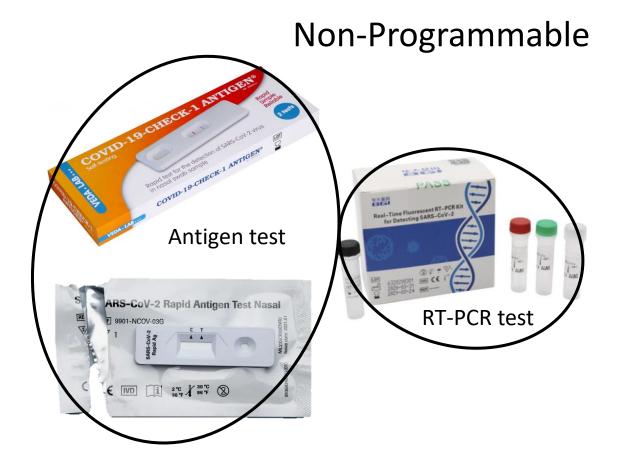




#### Programmable



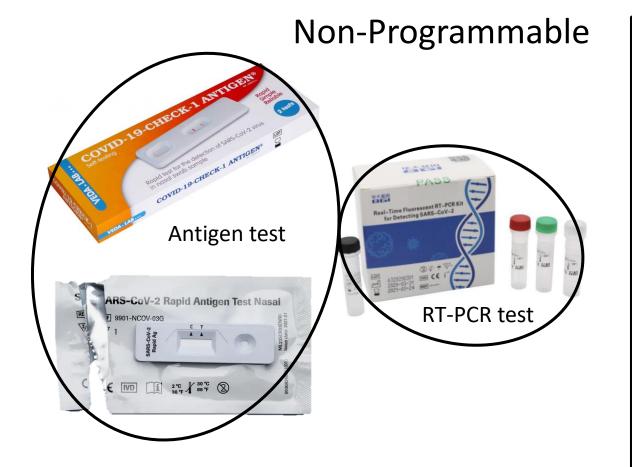




Programmable













#### Motivation

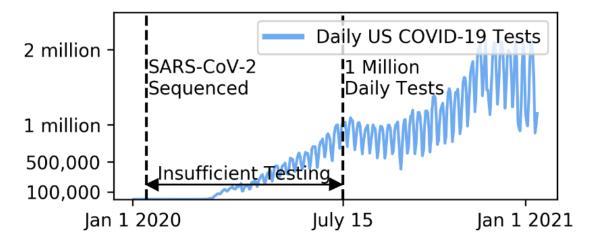


Figure 2: Progression of US COVID-19 testing [15]

#### Motivation

Non-programmable virus detectors are slow to deploy.

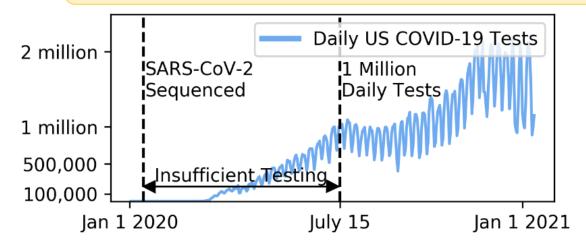


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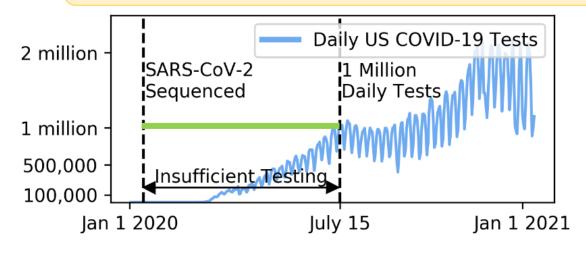


Figure 2: Progression of US COVID-19 testing [15]

**Programmable** virus detectors can help bridge the gap early in a pandemic.



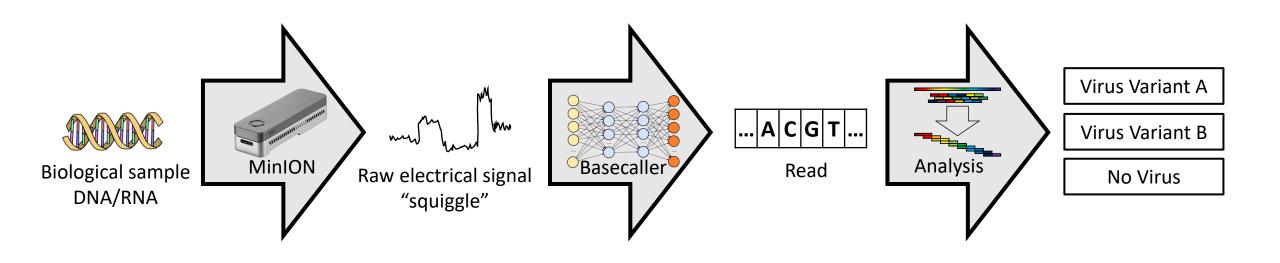


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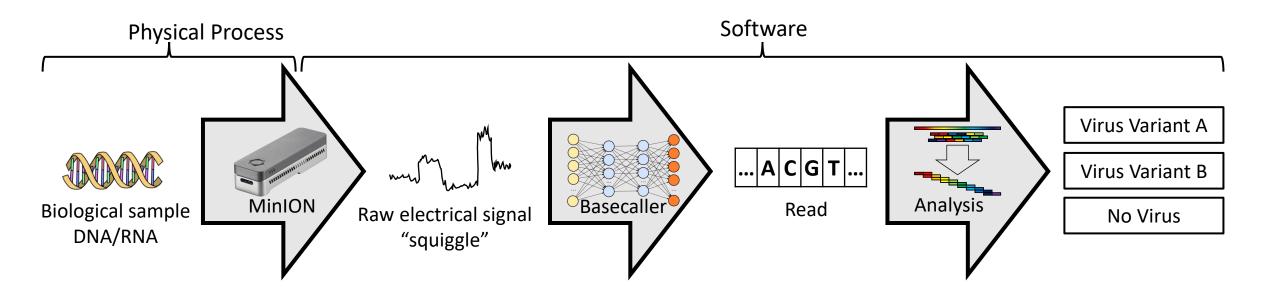






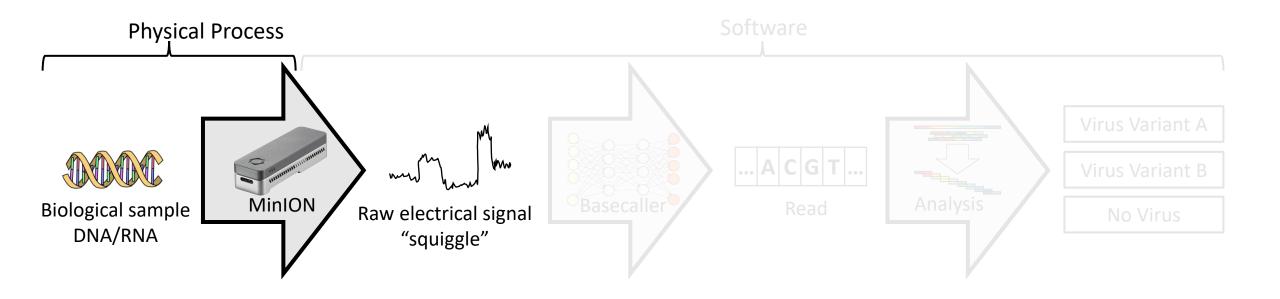






















#### MinION is

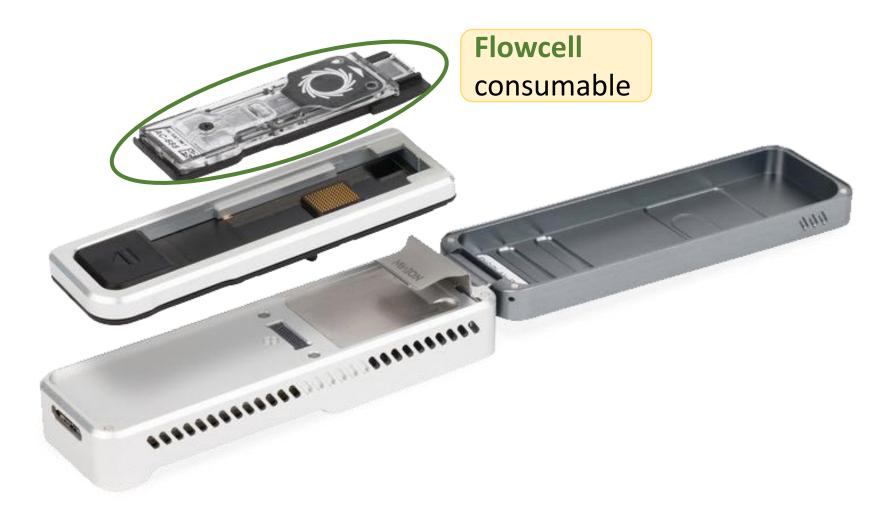
- cheap (\$1000 + \$hundreds for consumables)
- portable
- and supports ReadUntil



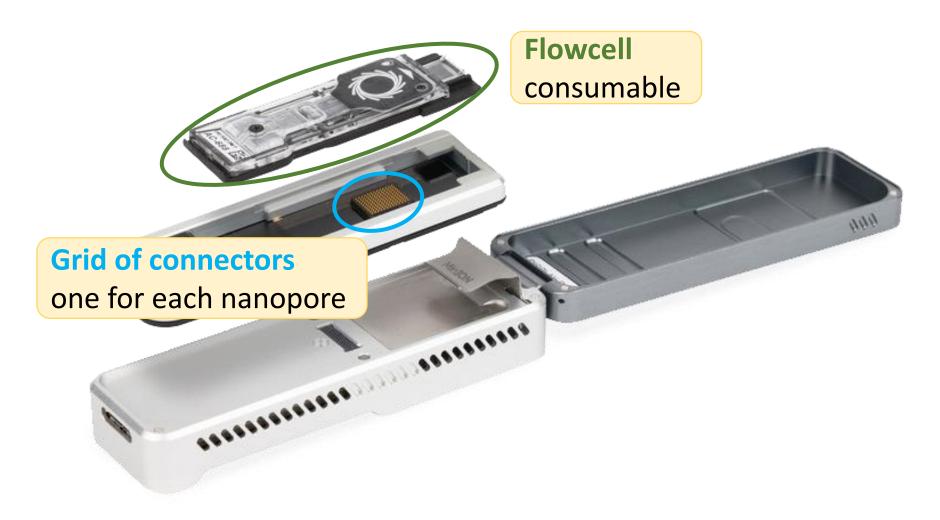






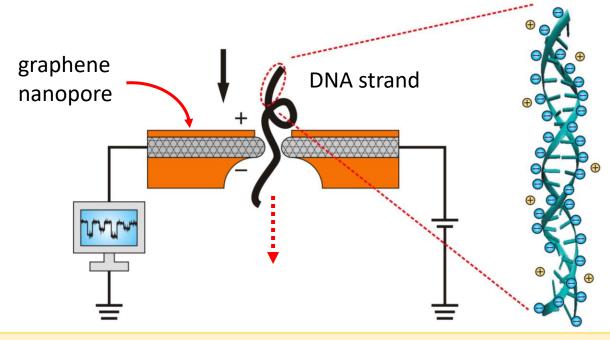








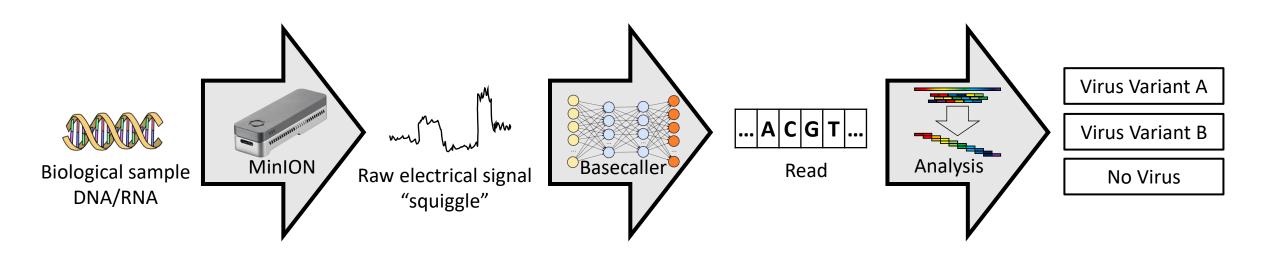
### How Do Nanopores Work?



- Nanopore is a nano-scale hole (<20nm).
- In nanopore sequencers, an **ionic current** passes through the nanopores
- When the DNA strand passes through the nanopore, the sequencer measures the change in current
- This change is used to identify the bases in the strand with the help of different electrochemical structures of the different bases

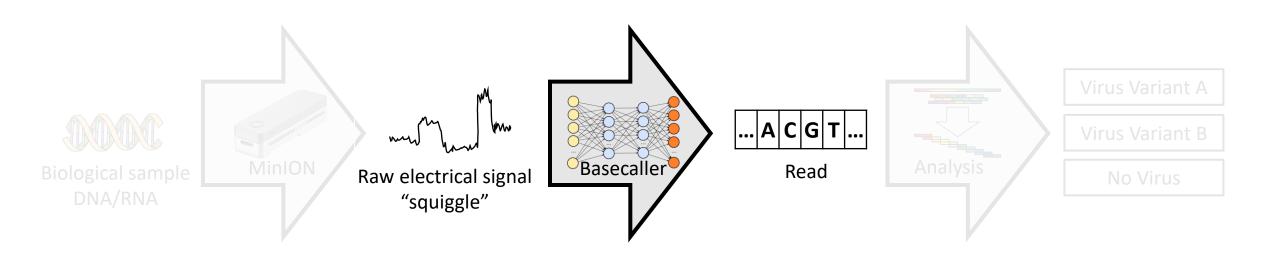








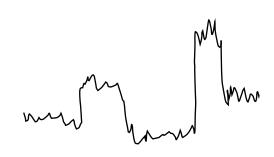




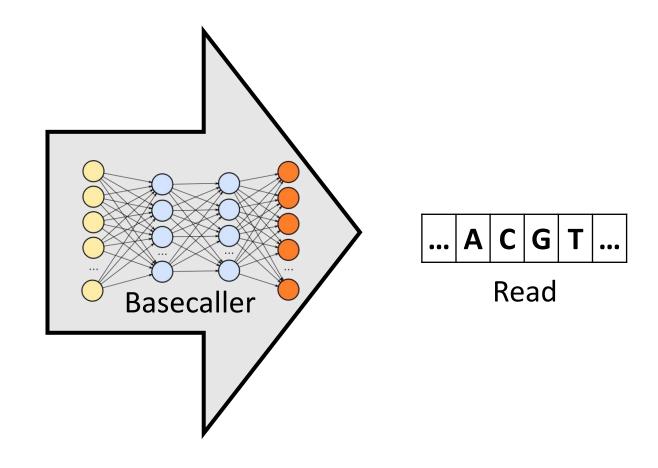




## Basecalling



Raw electrical signal "squiggle"



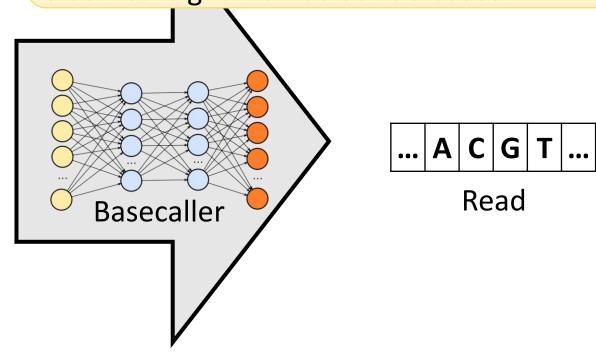


### Basecalling

my my my

Raw electrical signal "squiggle"

State-of-the-art basecallers use computationally expensive **deep neural networks** to convert the electrical signal to nucleotide bases.





### Basecalling Cost

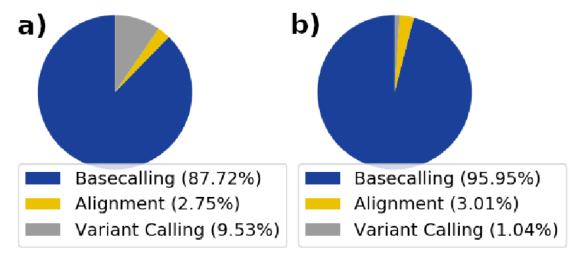


Figure 5: Basecalling is the bottleneck in a Read Until assembly of a SARS-CoV2 genome from specimens with a) 1%, and b) 0.1% viral reads.

### Basecalling Cost

Basecalling is the **sole bottleneck** of the computational steps.

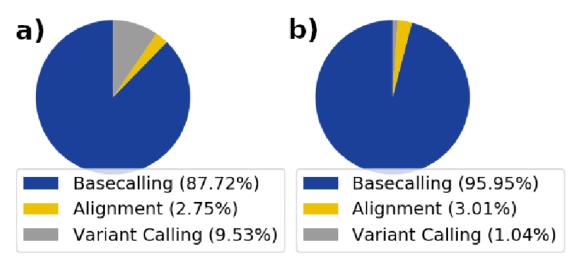
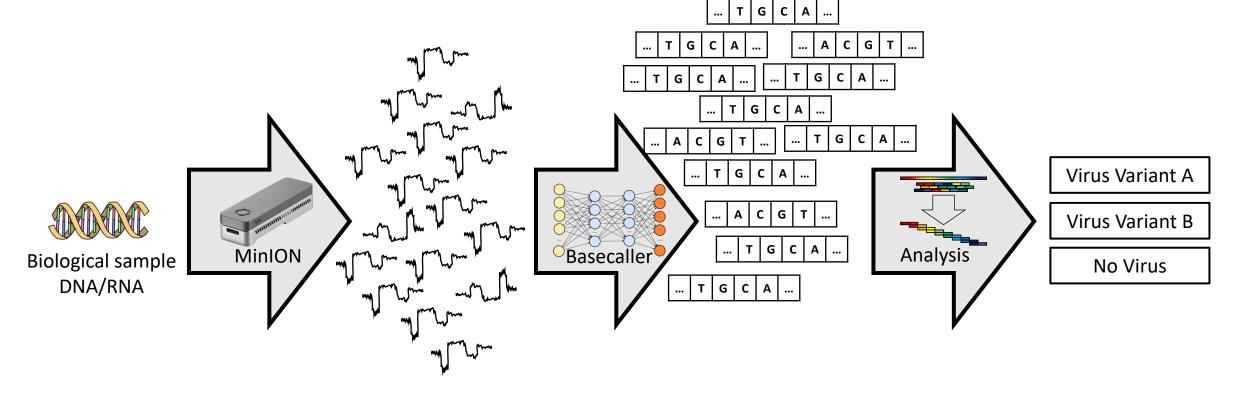


Figure 5: Basecalling is the bottleneck in a Read Until assembly of a SARS-CoV2 genome from specimens with a) 1%, and b) 0.1% viral reads.



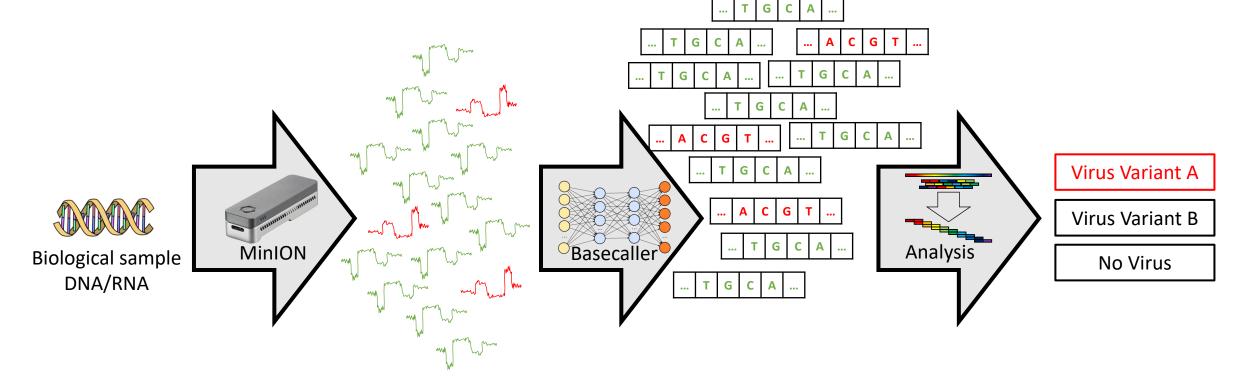
## Required Squiggles







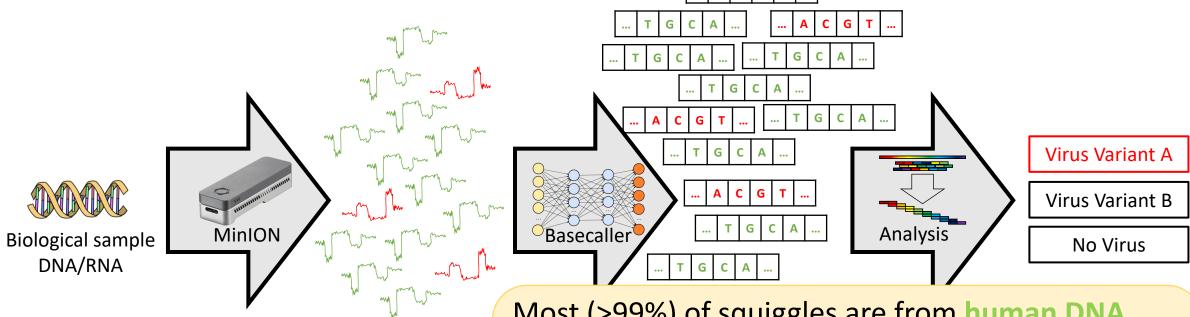
## Required Squiggles







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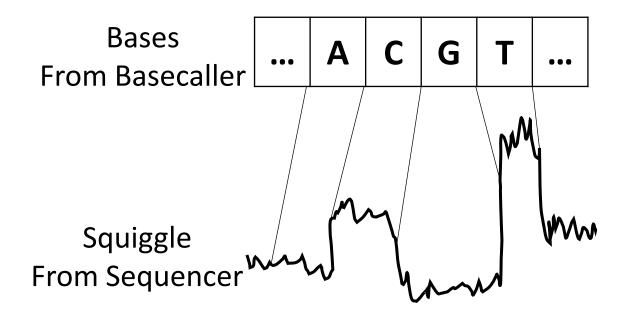
Most (>99%) of squiggles are from human DNA. However, only in viral squiggles are of interest for virus detection.

Thus, most of the basecaller's work is unnecessary.



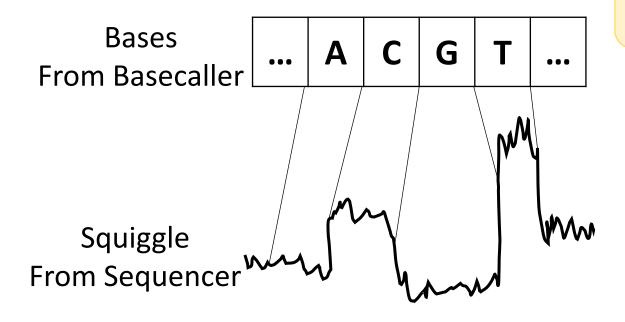


### Without ReadUntil





### Without ReadUntil

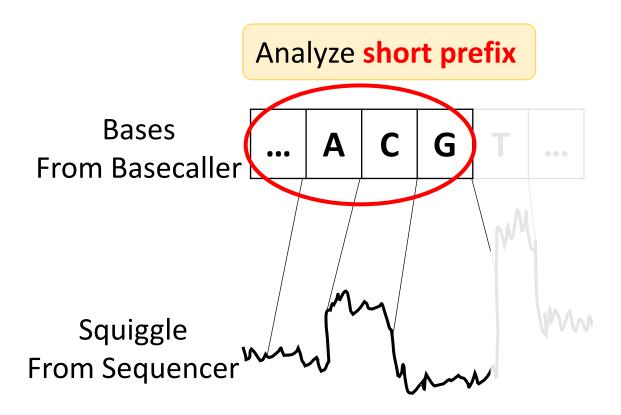


Without ReadUntil, the **entire squiggle** is processed by the machine.





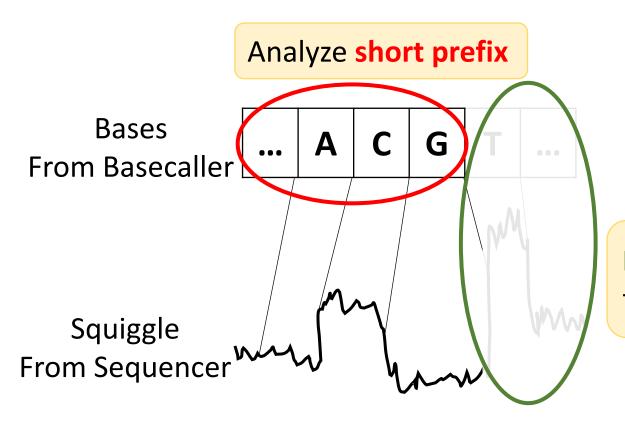
### Without ReadUntil







#### With ReadUntil

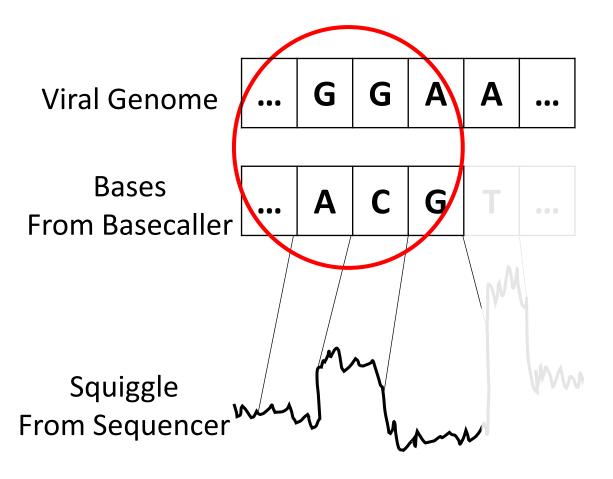


Remaining bases are not read if the prefix looks unpromising





#### With ReadUntil

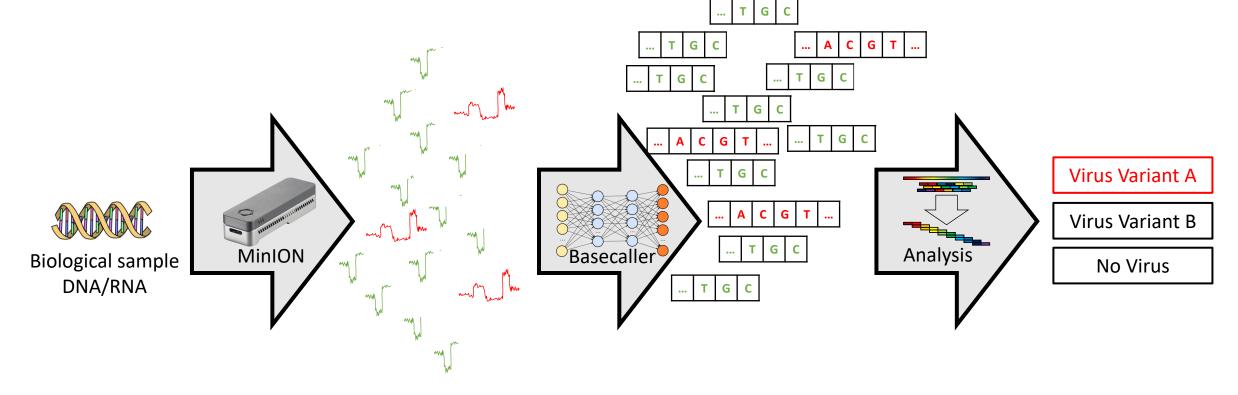


For example, compare the prefix to the virus genome we are looking for





## Required Squiggles







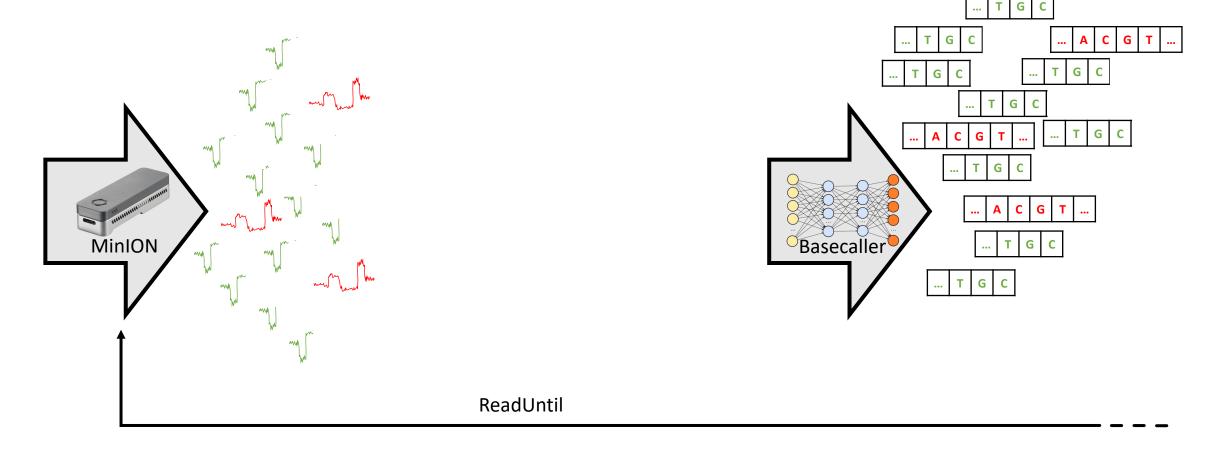
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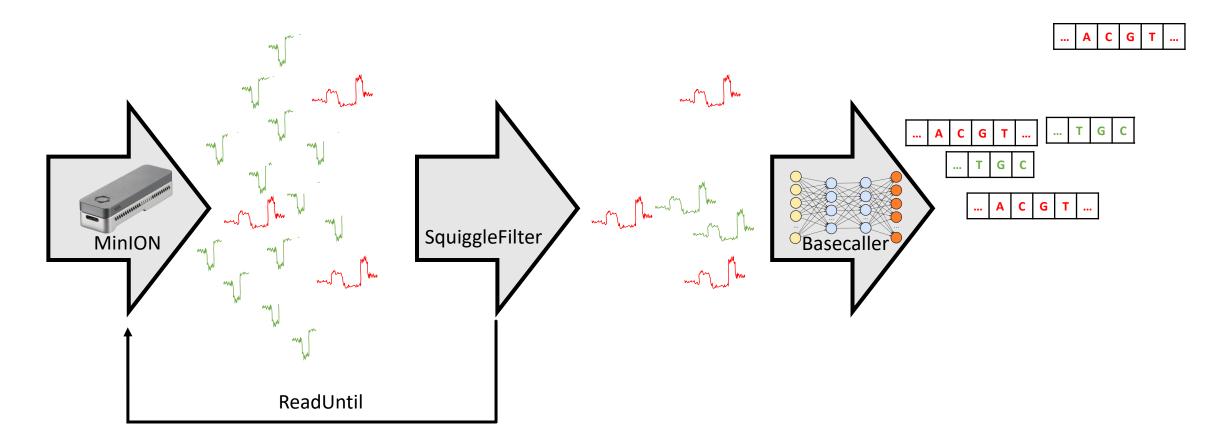
# Before SquiggleFilter







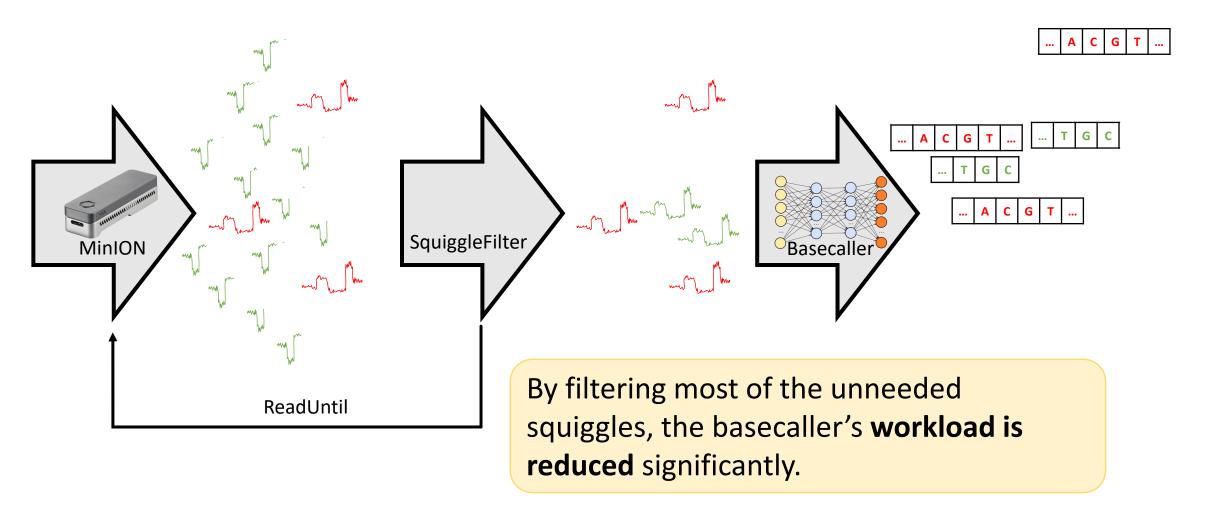
# With SquiggleFilter







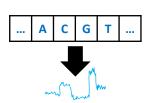
### With SquiggleFilter





# SquiggleFilter

1. Translate reference genome to expected squiggle



- 3. Compare input and reference signal using dynamic time warping

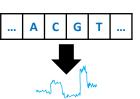


4. If they match: keep analyzing If not: reject immediately



## SquiggleFilter

1. Translate reference genome to expected squiggle



- 2. Normalize query squiggle
- 3. Compare input and reference signal using dynamic time warping
- 4. If they match: keep analyzing If not: reject immediately





## Expected Squiggle

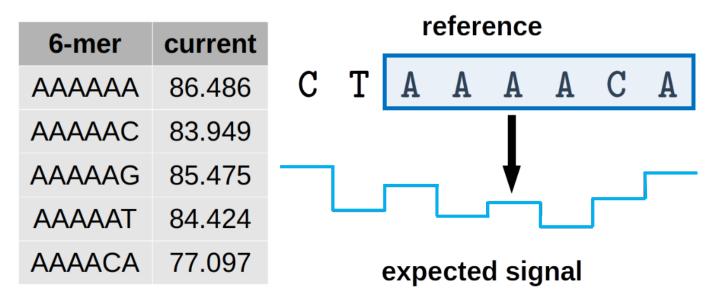


Figure 7: Aligning reference bases to expected currents.



## Expected Squiggle

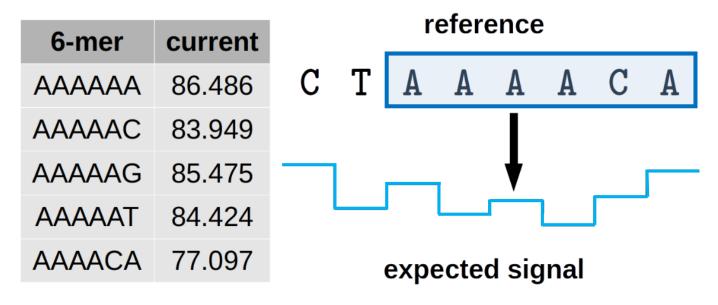


Figure 7: Aligning reference bases to expected currents.

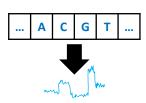
Each reference 6-mer corresponds to a current level, the mapping is provided by the manufacturer. The expected signal can be obtained through lookups from the table.





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# SquiggleFilter

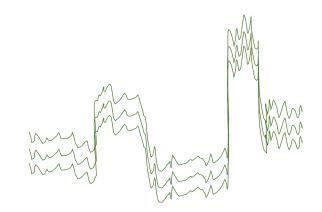
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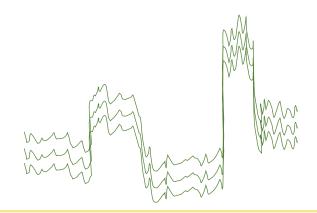








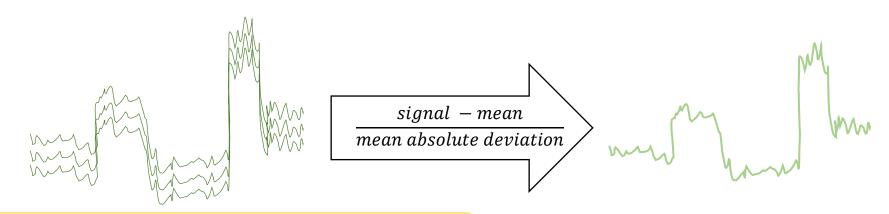




Measured currents in query squiggles can vary due to process variation, among other things.





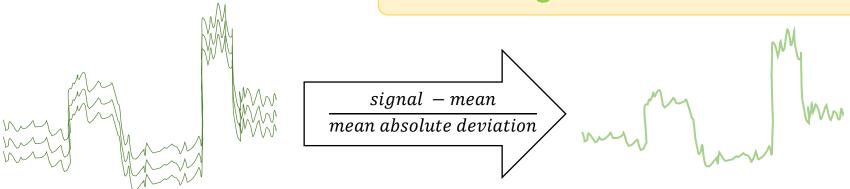


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Normalizing removes these differences.

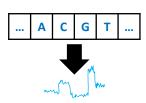


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### Dynamic Time Warping (prior work)



Vintsyuk, T.K. <u>Speech discrimination by dynamic programming</u>. *Cybernetics and System Analysis* **4,** 52–57 (1968).



### Dynamic Time Warping (prior work)

Reference Squiggle



Query squiggles are stretched on the x-axis, relative to the reference squiggle.

Query Squiggle



Vintsyuk, T.K. <u>Speech discrimination by dynamic programming</u>. *Cybernetics and System Analysis* **4,** 52–57 (1968).



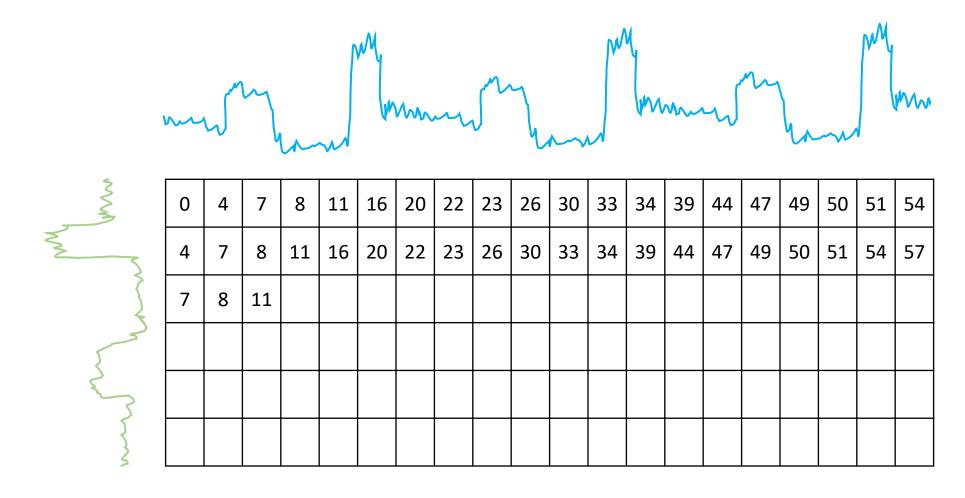
### Dynamic Time Warping (prior work)

Query squiggles are stretched on the xaxis, relative to the reference squiggle. Dynamic time warping finds the stretch Reference Squiggle that matches the squiggles best. **Query Squiggle** 

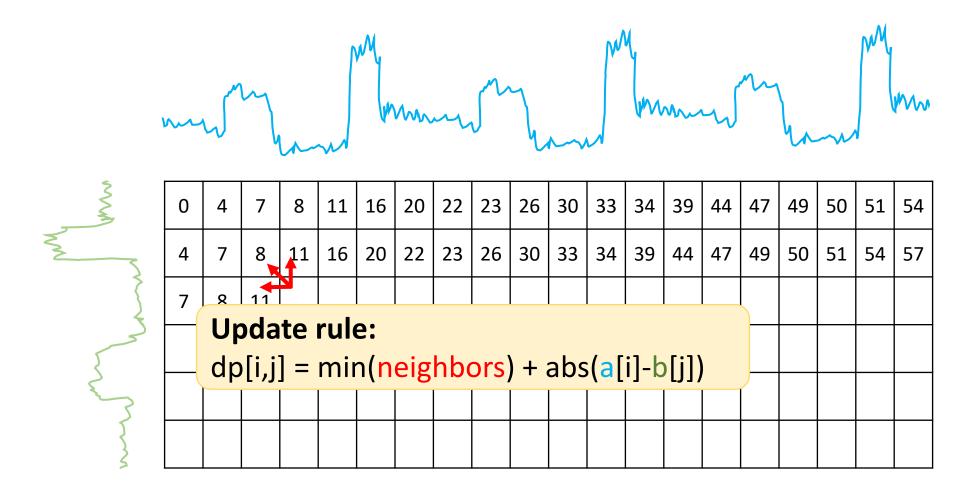
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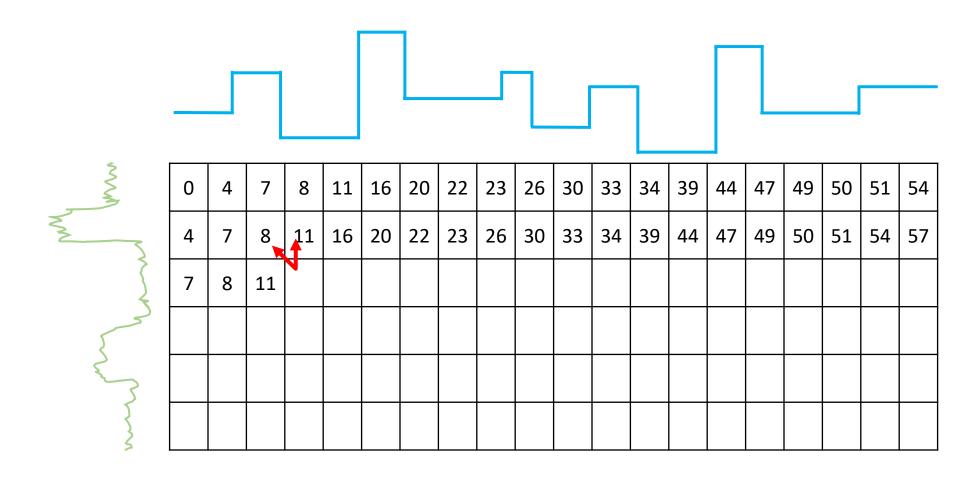






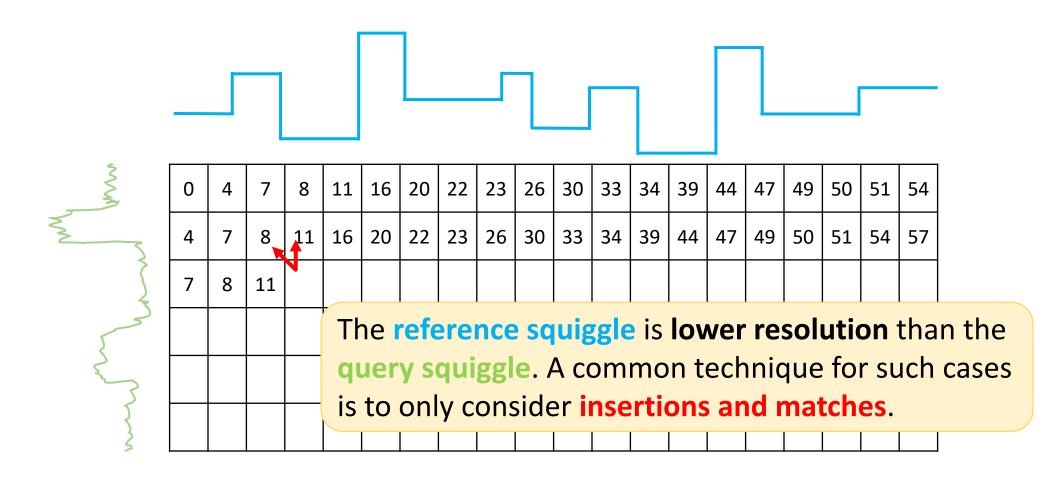






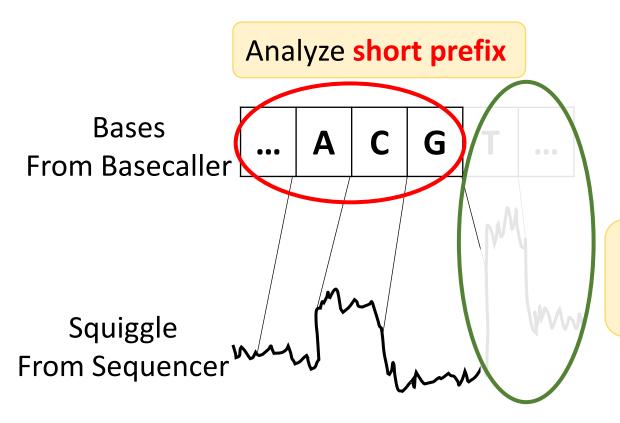








#### With ReadUntil

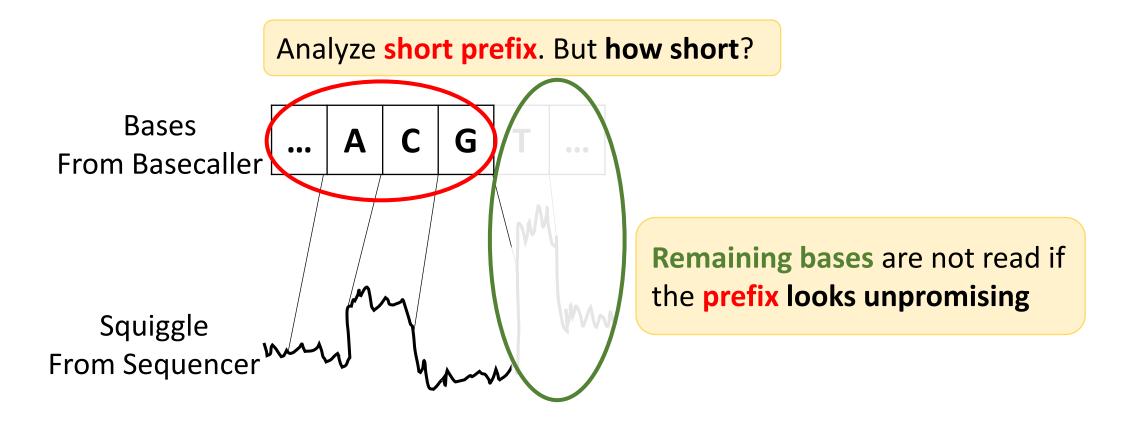


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#### With ReadUntil





# Query Prefix Size

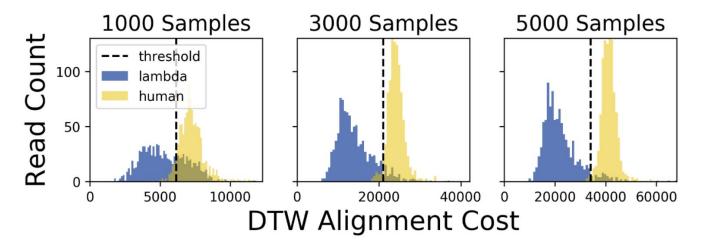


Figure 11: sDTW cost distributions for reads of 3 prefix lengths, aligned to the lambda phage genome.





# Query Prefix Size

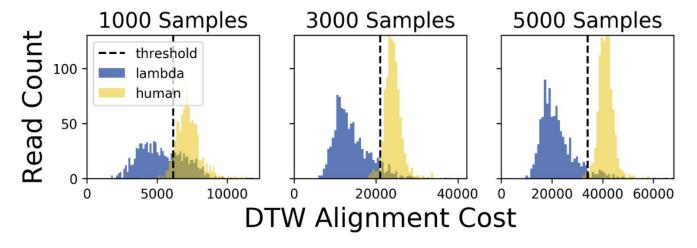
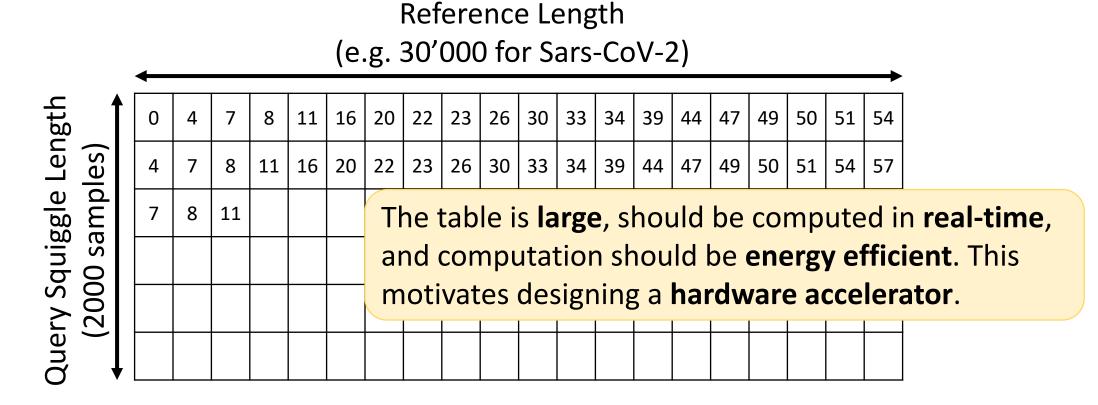


Figure 11: syllengths, alig

The authors determine **2000 samples** to be a good **middle ground** between accuracy and performance.









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#### **Accelerator Overview**

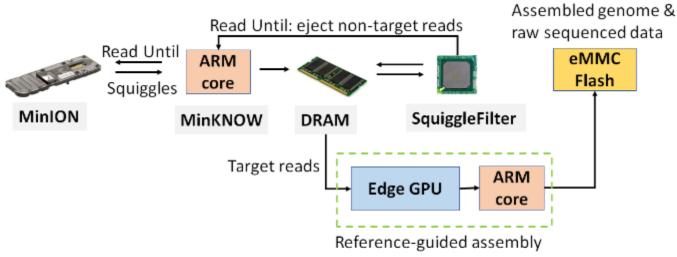


Figure 12: System-on-Chip design with the accelerated hardware filter on ASIC integrated with NVIDIA GPU and 8-core ARM v8.2 64-bit CPU



## Accelerator Systolic Array

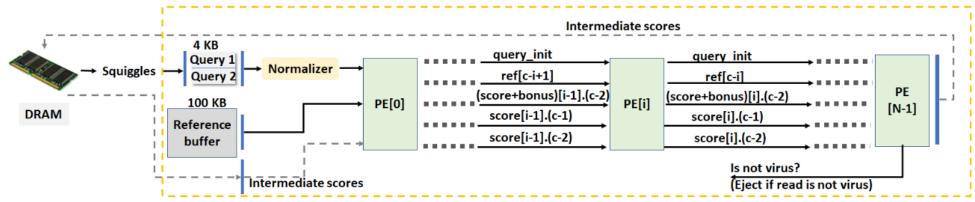


Figure 13: SquiggleFilter Tile. N=2000 PEs are connected with streaming inputs and outputs. The last PE determines the classification by comparing its cost to a threshold every cycle. c is the cycle and i is the PE index.



## Accelerator Systolic Array

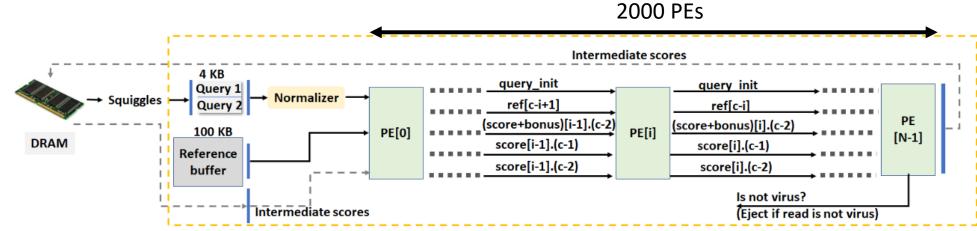


Figure 13: SquiggleFilter Tile. N=2000 PEs are connected with streaming inputs and outputs. The last PE determines the classification by comparing its cost to a threshold every cycle. c is the cycle and i is the PE index.

The DTW accelerator is implemented as a systolic array of 2000 PEs.





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### Results – Computational Speedup

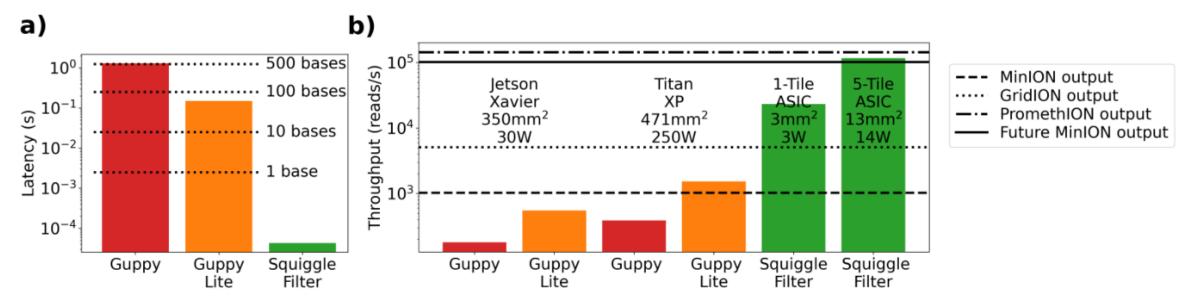


Figure 16: a) Latency, and b) throughput of Guppy, Guppy-lite and SquiggleFilter during Read Until.



### Results - Computational Speedup

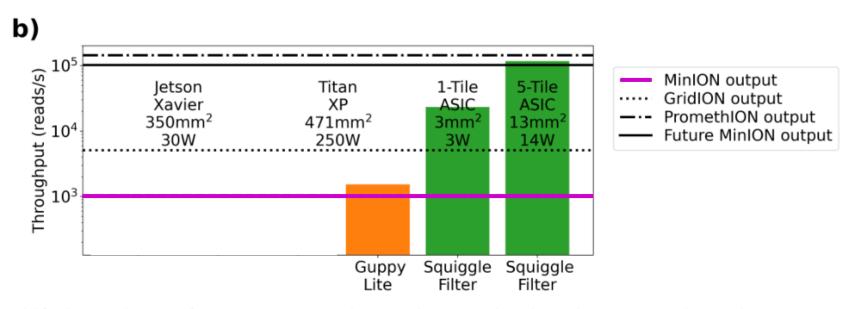


Figure 16: a) Latency, and b) throughput of Guppy, Guppy-lite and SquiggleFilter during Read Until.

Only the most inaccurate model on a server-class GPU or SquiggleFilter can keep up with even the slowest sequencer.





### Results - Computational Speedup

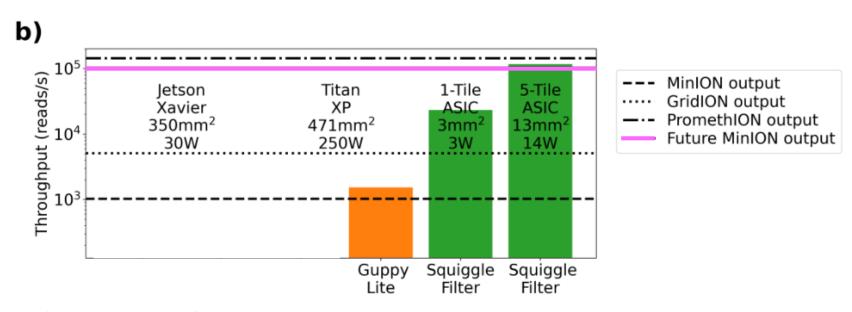
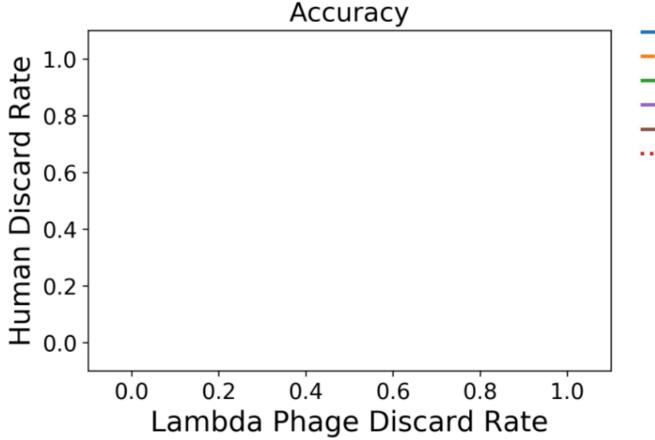


Figure 16: a) Latency, and b) throughput of Guppy, Guppy-lite and SquiggleFilter during Read Until.

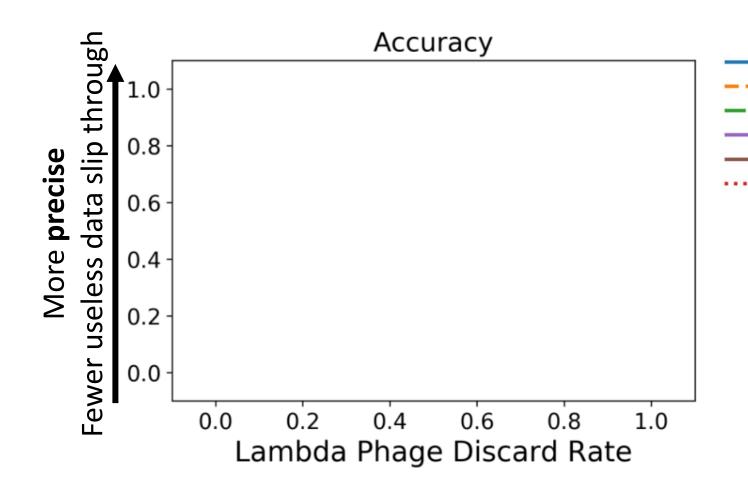
Only **SquiggleFilter** can keep up with a projected future version of the slowest sequencer.







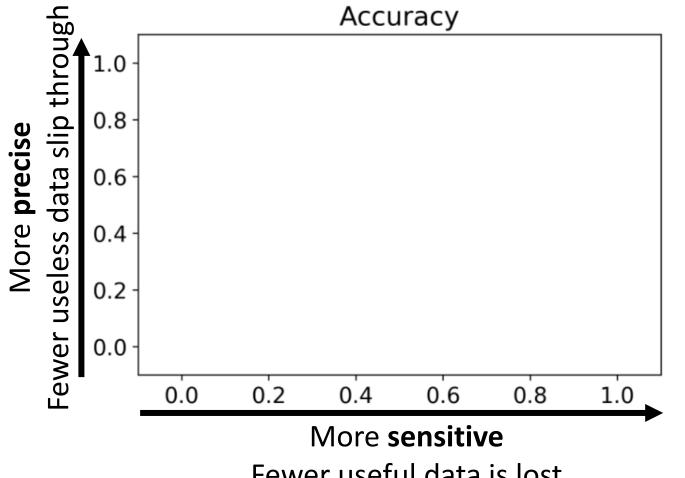






SquiggleFilter 1000 samples



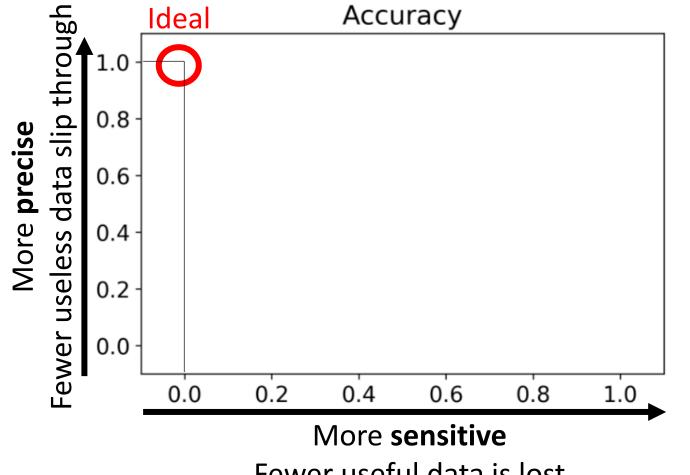


SquiggleFilter 1000 samples SquiggleFilter 2000 samples SquiggleFilter 3000 samples SquiggleFilter 5000 samples SquiggleFilter 8000 samples Guppy-lite 1000 samples

Fewer useful data is lost







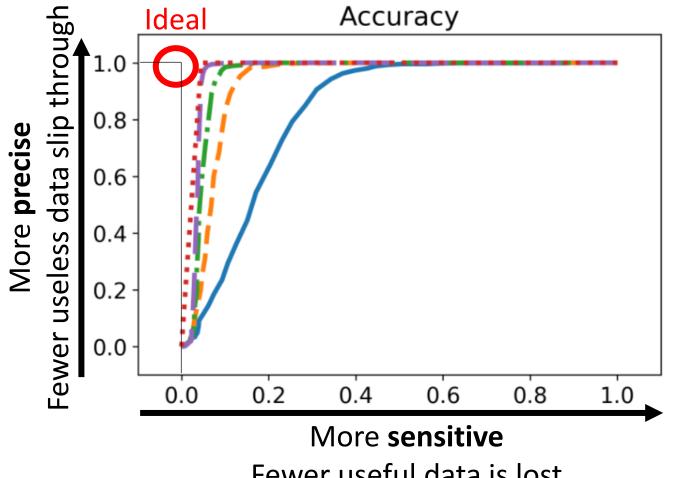


SquiggleFilter 1000 samples SquiggleFilter 2000 samples

SquiggleFilter 3000 samples SquiggleFilter 5000 samples

SquiggleFilter 8000 samples

Guppy-lite 1000 samples



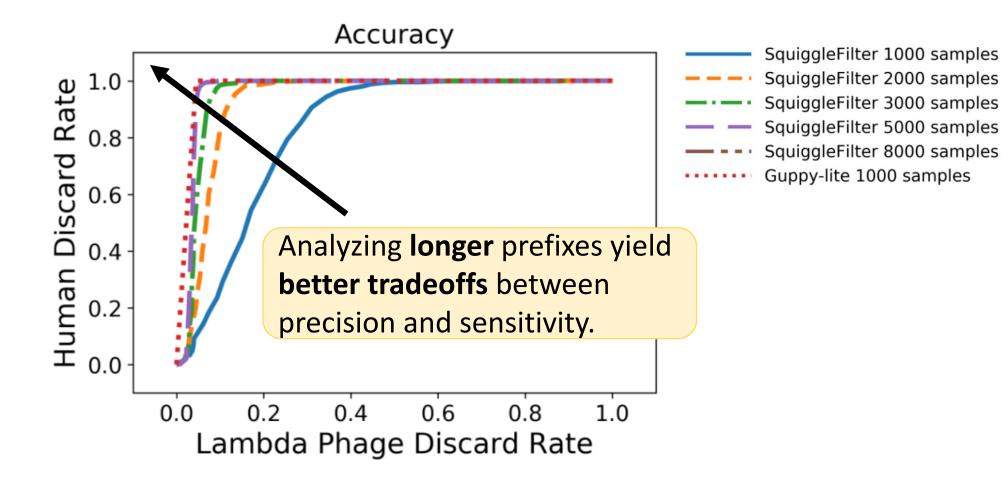


SquiggleFilter 1000 samples SquiggleFilter 2000 samples SquiggleFilter 3000 samples SquiggleFilter 5000 samples

SquiggleFilter 8000 samples

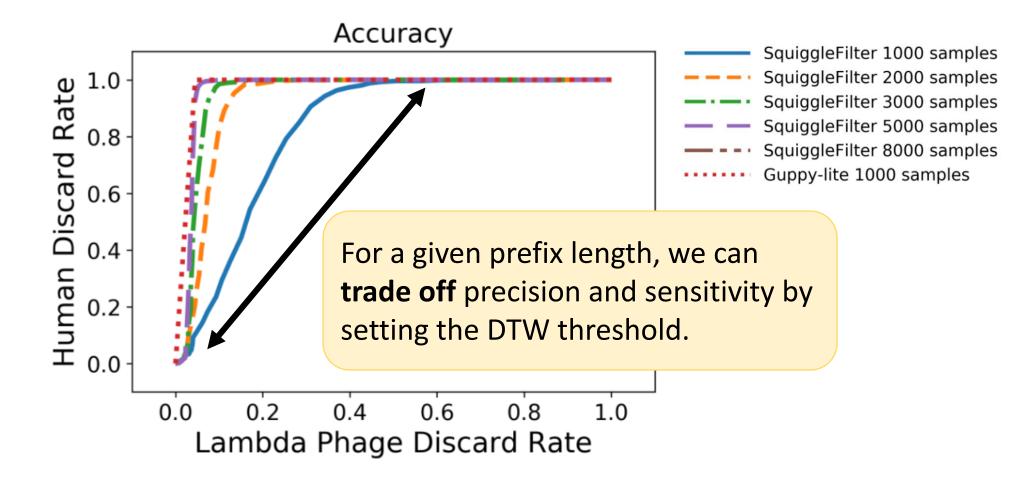
Guppy-lite 1000 samples

Fewer useful data is lost



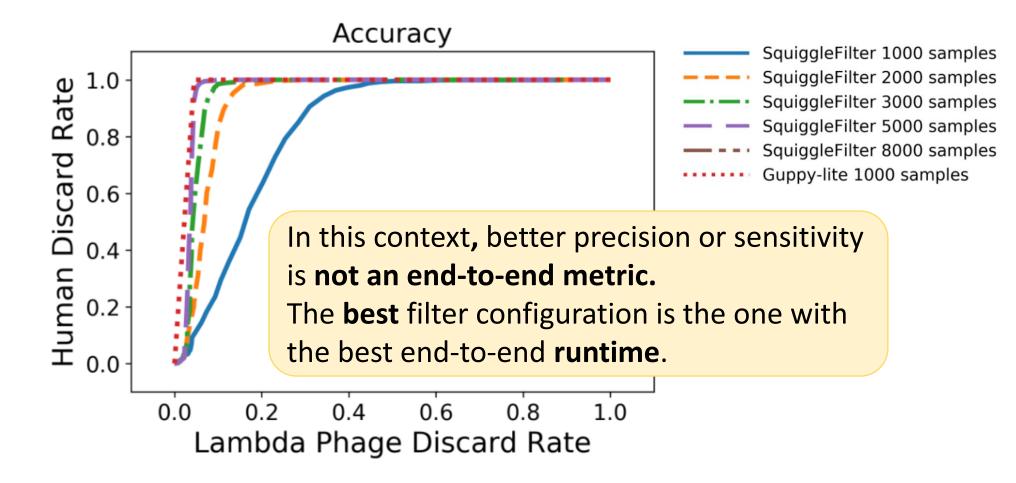






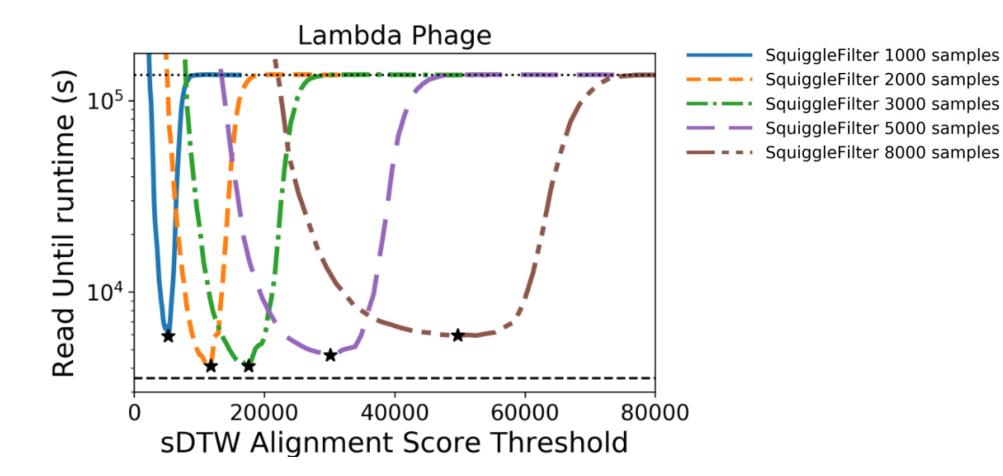






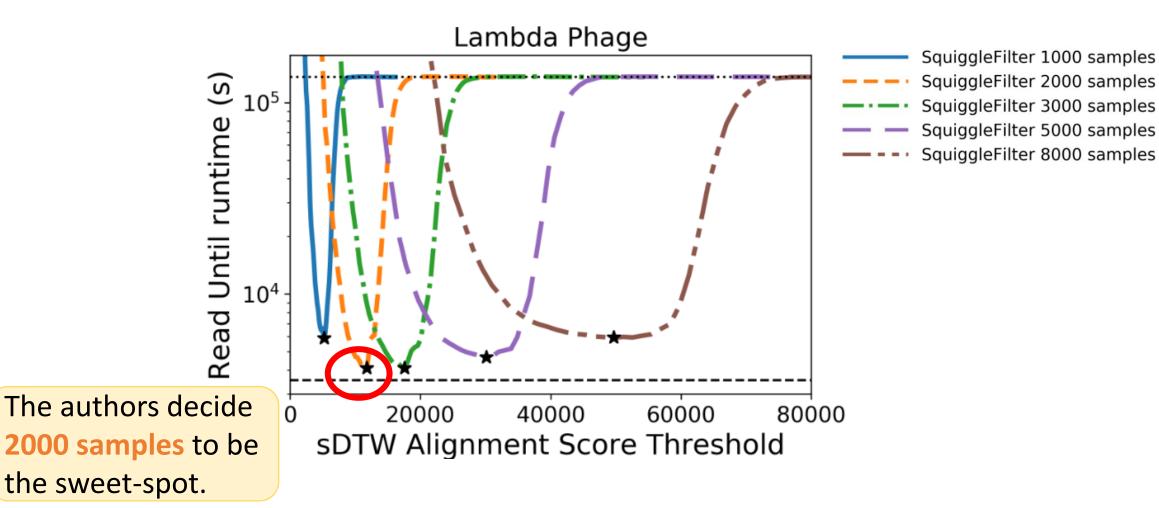






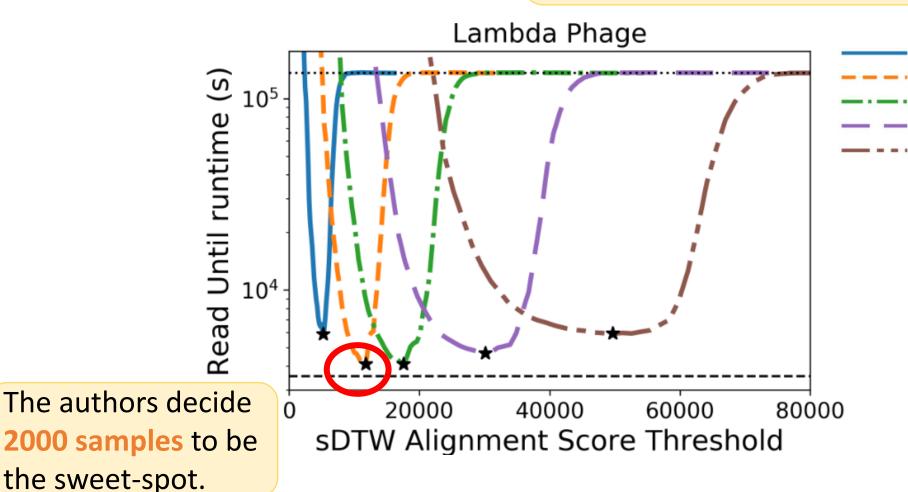








SquiggleFilter achieves an **end-to-end** speedup over Guppy-lite of **12.9%**. (Not drawn here)





SquiggleFilter 1000 samples

SquiggleFilter 2000 samples

SquiggleFilter 3000 samples

SquiggleFilter 5000 samples SquiggleFilter 8000 samples

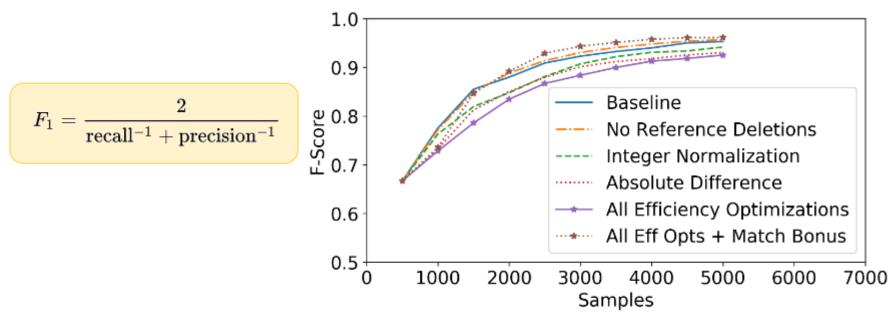


Figure 18: Accuracy results for modifications to the standard sDTW algorithm.



#### Results - Power

<b>ASIC Element</b>	Area (mm²)	Power (W)
Normalizer	0.014	0.045
<b>Processing Element</b>	0.001	0.002
Tile (1×2000 PEs)	2.423	2.780
Query buffer	0.023	0.009
Reference buffer	0.185	0.028
Complete 1-Tile ASIC Complete 5-Tile ASIC	2.65 13.25	2.86 14.31

Table 4: SquiggleFilter ASIC synthesis results.

Synthesized for **28nm** TSMC HPC @2.5GHz



#### Results - Power

SquiggleFilter's **ASIC** is significantly more **power efficient than** a server-class GPU, sufficiently so it could be battery powered.

ASIC Element	Area (mm²)	Power (W)
Normalizer	0.014	0.045
<b>Processing Element</b>	0.001	0.002
Tile (1×2000 PEs)	2.423	2.780
Query buffer	0.023	0.009
Reference buffer	0.185	0.028
Complete 1-Tile ASIC Complete 5-Tile ASIC	2.65 13.25	2.86 14.31

Table 4: SquiggleFilter ASIC synthesis results.

Synthesized for **28nm** TSMC HPC @2.5GHz





#### Outline

- 1. Background and Motivation
  - 1. Virus detection
  - 2. Nanopore sequencing
- 2. SquiggleFilter
  - 1. Pipeline
  - 2. Accelerator
- 3. Results
- 4. Strengths/Weaknesses
- 5. Discussion





• Extremely **important** problem



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- The solution works well and is reasonable to implement





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- Future proof (can tolerate much faster sequencers)





- Extremely important problem
- The solution works well and is reasonable to implement
- Future proof (can tolerate much faster sequencers)
- They claim to be the first proposal of using squiggle alignment for enriching low-concentration viral specimen with ReadUntil



• **No comparison to a CPU/GPU** implementation of their algorithm. It's unclear how much of the speedup comes from the algorithm, and how much from HW acceleration





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- No comparison to a CPU/GPU implementation of their algorithm. It's unclear how much of the speedup comes from the algorithm, and how much from HW acceleration
- No comparison to other DTW accelerators
- No coverage accuracy measurements
- The work is limited to relatively small viral genomes. Their proposal would not work for viruses with a larger genome, such as Smallpox and Herpes Simplex





#### Weakness – Limited to Small Genomes

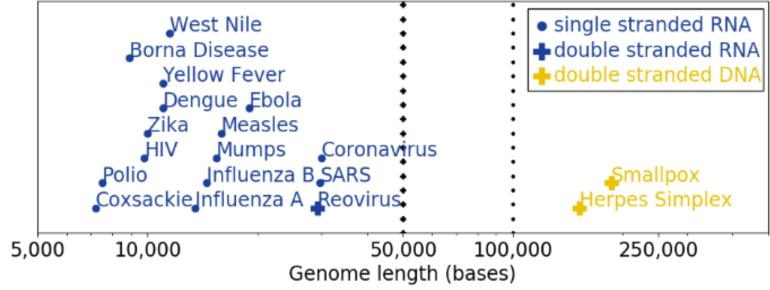


Figure 10: Epidemic virus genome lengths.





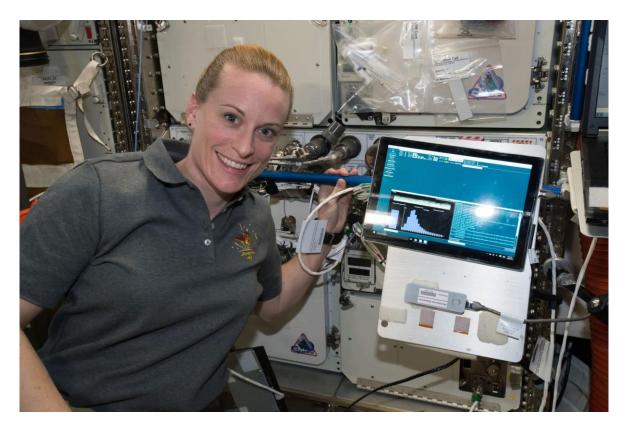
#### Outline

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#### What are challenges of genome analysis in space?



https://www.universetoday.com/135327/whats-strange-glowing-mold-astronauts-will-soon-able-sequence-unknown-space-organisms/





#### What are challenges of genome analysis in space?

Wallace offered support to Whitson, a biochemist, as she used the MinION device (developed by Oxford Nanopore Technologies) to sequence the amplified DNA.

The data were downlinked to the team in Houston for analysis and identification.

"Once we actually got the data on the ground we were able to turn it around and start analyzing it," said Aaron Burton, NASA biochemist and the project's co-investigator. "You get all these **squiggle plots** and you have to turn that into As, Gs, Cs and Ts."

https://www.nasa.gov/mission\_pages/station/research/news/b4h-3rd/ge-gis-3-identifies-unknown-microbes





#### What are challenges of genome analysis in space?

- Low communication bandwidth with base station
- Devices must be low energy, lightweight, portable
- Devices should be reconfigurable









• Fourier Transform, then compare in frequency space?





- Fourier Transform, then compare in frequency space?
- Compare based on handcrafted metrics?
  - Number of peaks and valleys
  - Bin oscillations and compare distributions
  - ...





- Fourier Transform, then compare in frequency space?
- Compare based on handcrafted metrics?
  - Number of peaks and valleys
  - Bin oscillations and compare distributions
  - ...
- Fortunately, being inaccurate only costs time, so we are free to experiment wildly in this application!





# Can we apply heuristics to DTW?





#### Can we apply heuristics to DTW?

- Search less entries of the O(n\*m) matrix
- Several techniques exist for the (related) approximate string matching problem
  - Banded diagonals, such as when calculating edit distance <u>Algorithms for approximate string matching</u>, E. Ukkonen, 1985
  - Greedy
     GenASM: A High-Performance, Low-Power Approximate String Matching Acceleration Framework for Genome Sequence Analysis, Senol Cali et al., 2020
  - Seeding

    <u>Targeted nanopore sequencing by real-time mapping of raw electrical signal with UNCALLED</u>, Kovaka et al., 2021





# SquiggleFilter

### An Accelerator for Portable Virus Detection

Tim Dunn, Harisankar Sadasivan, Jack Wadden, Kush Goliya, Kuan-Yu Chen, David Blaauw, Reetuparna Das, and Satish Narayanasamy University of Michgan MICRO '21

Presented by: Joël Lindegger joel.lindegger@inf.ethz.ch 23/12/2021





# Backup Slides







Non-Programmable









Non-Programmable

- Tailor chemistry to the given virus
- Cheaply mass-manufacture chemistry
- Kits must be physically distributed









#### Non-Programmable

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- Distribute viral genome (Software)
- Sequence DNA/RNA reads
- Compare to viral genome (Software)







Non-Programmable

- Tailor chemistry to the given virus
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Non-Programmable

- Tailor chemistry to the given virus
- Cheaply mass-manufacture chemistry
- Kits must be physically distributed
- Inflexible



- Distribute viral genome (Software)
- Sequence DNA/RNA reads
- Compare to viral genome (Software)
- Flexible







#### Non-Programmable

- Tailor chemistry to the given virus
- Cheaply mass-manufacture chemistry
- Kits must be physically distributed
- Inflexible
- Unavailable at beginning of pandemic



- **Distribute viral genome (Software)**
- Sequence DNA/RNA reads
- Compare to viral genome (Software)
- Flexible
- Available immediately







#### Non-Programmable

- Tailor chemistry to the given virus
- Cheaply mass-manufacture chemistry
- Kits must be physically distributed
- Inflexible
- Unavailable at beginning of pandemic
- Low diagnostic power (virus/no virus)





- Distribute viral genome (Software)
- Sequence DNA/RNA reads
- Compare to viral genome (Software)
- Flexible
- Available immediately
- High diagnostic power (e.g. virus strain)







#### Non-Programmable

- Tailor chemistry to the given virus
- Cheaply mass-manufacture chemistry
- Kits must be physically distributed
- Inflexible
- Unavailable at beginning of pandemic
- Low diagnostic power (virus/no virus)
- Cheap





- Distribute viral genome (Software)
- Sequence DNA/RNA reads
- Compare to viral genome (Software)
- Flexible
- Available immediately
- High diagnostic power (e.g. virus strain)
- Expensive







#### Non-Programmable

- Tailor chemistry to the given virus
- Cheaply mass-manufacture chemistry
- Kits must be physically distributed
- Inflexible
- Unavailable at beginning of pandemic
- Low diagnostic power (virus/no virus)
- Cheap
- Fast





- Distribute viral genome (Software)
- Sequence DNA/RNA reads
- Compare to viral genome (Software)
- Flexible
- Available immediately
- High diagnostic power (e.g. virus strain)
- Expensive
- Slow





# Related work





# Do signal comparison algorithms/accelerators from other domains already exist?

# NATSA: A Near-Data Processing Accelerator for Time Series Analysis

Ivan Fernandez§	Ricardo Quislant <sup>§</sup>		Christina Giannoula <sup>†</sup>		Mohammed Alser <sup>‡</sup>
Juan Gómez-L	una <sup>‡</sup>	Eladio Gutié	errez§	Oscar Plata <sup>§</sup>	Onur Mutlu <sup>‡</sup>
§University of Mal	aga	$^{\dagger}National$	Technical	University of Athens	<sup>‡</sup> ETH Zürich

Time series analysis is a key technique for extracting and predicting events in domains as diverse as epidemiology, genomics, neuroscience, environmental sciences, economics, and more. *Matrix profile*, the state-of-the-art algorithm to perform time series analysis, computes the most similar subsequence for a given query subsequence within a sliced time series. *Matrix profile* has low arithmetic intensity, but it typically operates on large amounts of time series data. In current computing systems, this data needs to be moved between the off-chip memory units and the on-chip computation units for

tection, which cannot be tolerated by many applications (e.g., vehicle safety systems [85]). Unlike approximate algorithms, exact algorithms [67] do not yield false positives or discordant dismissals, but can be very time-consuming on large time series data. Thus, *anytime* versions (aka interruptible algorithms) of exact algorithms are proposed to provide approximate solutions quickly [108, 112] and can return a valid result even if the user stops their execution early.

The state-of-the-art exact *anytime* method for motif and discord discovery is *matrix profile* [108], which is based on Eu-

ICCD 2020, pp. 120-129





# Do raw signals contain extra information?

# Trends in **Genetics**



#### Review

Beyond sequencing: machine learning algorithms extract biology hidden in Nanopore signal data

Yuk Kei Wan, 1,2 Christopher Hendra, 3,1 Ploy N. Pratanwanich, 1,4,5 and Jonathan Göke 1,6,\*

Nanopore sequencing provides signal data corresponding to the nucleotide motifs sequenced. Through machine learning-based methods, these signals are translated into long-read sequences that overcome the read size limit of short-read sequencing. However, analyzing the raw nanopore signal data provides many more opportunities beyond just sequencing genomes and transcriptomes: algorithms that use machine learning approaches to extract biological information from these signals allow the detection of DNA and RNA

#### Highlights

Nanopore sequencing accuracy has increased to 98.3% as new-generation base callers replace early generation hidden Markov model basecalling algorithms with neural network algorithms.

Machine learning methods can classify

https://www.cell.com/trends/genetics/fulltext/S0168-9525(21)00257-2





# Which sequencing technology is best for virus

detection?

nature biomedical engineering





# Massively scaled-up testing for SARS-CoV-2 RNA via next-generation sequencing of pooled and barcoded nasal and saliva samples

Joshua S. Bloom <sup>1,2,3</sup> <sup>1</sup>, Laila Sathe <sup>4</sup>, Chetan Munugala<sup>1,2</sup>, Eric M. Jones<sup>3</sup>, Molly Gasperini<sup>3</sup>, Nathan B. Lubock<sup>3</sup>, Fauna Yarza <sup>3</sup>, Erin M. Thompson<sup>3</sup>, Kyle M. Kovary<sup>3</sup>, Jimin Park<sup>3</sup>, Dawn Marquette <sup>5</sup>, Stephania Kay<sup>5</sup>, Mark Lucas<sup>5</sup>, TreQuan Love<sup>5</sup>, A. Sina Booeshaghi<sup>6</sup>, Oliver F. Brandenberg<sup>1,2,7</sup>, Longhua Guo<sup>1,2,7</sup>, James Boocock<sup>1,2,7</sup>, Myles Hochman<sup>3</sup>, Scott W. Simpkins <sup>3</sup>, Isabella Lin<sup>1,4</sup>, Nathan LaPierre<sup>8</sup>, Duke Hong<sup>5</sup>, Yi Zhang<sup>1</sup>, Gabriel Oland <sup>9</sup>, Bianca Judy Choe<sup>10</sup>, Sukantha Chandrasekaran<sup>4</sup>, Evann E. Hilt<sup>4</sup>, Manish J. Butte <sup>11,12</sup>, Robert Damoiseaux <sup>13,14,15</sup>, Clifford Kravit <sup>16</sup>, Aaron R. Cooper<sup>3</sup>, Yi Yin<sup>1</sup>, Lior Pachter <sup>17</sup>, Omai B. Garner<sup>4</sup>, Jonathan Flint <sup>1,18</sup>, Eleazar Eskin <sup>1,5,8</sup>, Chongyuan Luo<sup>1</sup>, Sriram Kosuri <sup>3,19</sup>, Leonid Kruglyak <sup>1,2,7</sup> and Valerie A. Arboleda <sup>1,4</sup>

Nature Biomedical Engineering 5, 657–665 (2021).





# Is early "viral"/"non-viral" labelling possible with Illumina as well?

